

Photobiomodulation for Neurodegenerative Diseases: A Scoping Review

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Abstract

Neurodegenerative diseases involve the progressive dysfunction and loss of neurons in the central nervous system and thus present a significant challenge due to the absence of effective therapies for halting or reversing their progression. Based on the characteristics of neurodegenerative diseases such as Alzheimer's disease (AD) and Parkinson's disease (PD), which have prolonged incubation periods and protracted courses, exploring non-invasive physical therapy methods is essential for alleviating

such diseases and ensuring that patients have an improved quality of life. Photobiomodulation (PBM) uses red and infrared light for therapeutic benefits and functions by stimulating, healing, regenerating, and protecting organizations at risk of injury, degradation, or death. Over the last two decades, PBM has gained widespread recognition as a non-invasive physical therapy method, showing efficacy in pain relief, anti-inflammatory responses, and tissue regeneration. Its application has expanded into the fields of neurology and psychiatry, where extensive research has been conducted. This paper presents a review and evaluation of studies investigating PBM in neurodegenerative diseases, with a specific emphasis on recent applications in AD and PD treatment for both animal and human subjects. Molecular mechanisms related to neuron damage and cognitive impairment are scrutinized, offering valuable insights into PBM's potential as a non-invasive therapeutic strategy.

Keywords: neurodegenerative diseases, photobiomodulation, Alzheimer's disease, Parkinson's disease

1. Introduction

Neurodegenerative diseases are a class of disorders characterized by the gradual degeneration and death of nerve cells (neurons) in the central nervous system (CNS) or

peripheral nervous system (PNS). This degeneration leads to a progressive decline in cognitive, motor, and/or sensory functions. Neurodegenerative diseases encompass a broad range of conditions, including Alzheimer's disease (AD), Parkinson's disease (PD), amyotrophic lateral sclerosis (ALS), Huntington's disease (HD), frontotemporal dementia (FTD), and prion disease. The global prevalence of neurodegenerative diseases is substantial and continues to increase due to the growing aging population. According to the World Alzheimer Report 2019 [1], approximately 50 million people worldwide have dementia, with AD being the most common cause. PD, ALS, and other neurodegenerative conditions collectively affect millions more across the globe [2,3]. Researchers are actively exploring genetic, environmental, and lifestyle factors that contribute to the increase in neurodegenerative diseases, aiming to develop effective treatments or interventions [4].

However, neurodegeneration is a physiological process that has yet to be completely fully understood. This paper reviews the preclinical and clinical evidence for the neurobiological and behavioral effects of a non-pharmacological physical therapy method, photobiomodulation (PBM) therapy, in brain degeneration and proposes the necessity of more controlled studies.

2. Treatment of Neurodegenerative

Diseases

Currently, researchers and doctors are actively exploring various treatment methods, including drug therapy, gene therapy, immunotherapy, and cell therapy, to address neurodegenerative diseases.

Drug therapy: For Alzheimer's disease, drug therapies such as cholinesterase inhibitors (e.g., donepezil) are used to improve cognitive function [5]. Hormone therapy and antidepressants are also employed for symptom management. For PD, dopamine replacement therapy, mainly with levodopa, is the primary treatment, and other medications (e.g., monoamine oxidase type B inhibitors, amantadine, anticholinergics, β -blockers, or dopamine agonists) may be initiated first to avoid levodopa-related motor complications [6]. Deep brain stimulation and medication adjustments are commonly used for symptom relief.

Gene therapy: For Alzheimer's disease, gene-based therapeutic targets include APP, MAPT, and APOE [7]. For Huntington's disease, researchers are developing gene-editing techniques targeting the Huntington's disease gene mutation to alleviate disease symptoms [8].

Immunotherapy: For Alzheimer's disease, some studies focus on using antibodies or vaccines to clear amyloid

plaques in the brain to slow disease progression [9].

Cell therapy: Scientists are actively developing cell-based therapies for diverse neurodegenerative diseases impacting the CNS. Small molecules and growth factors are employed to prompt human pluripotent stem cells (hPSCs) to generate neural progenitor cells (NPCs), resulting in specific lineages situated in distinct regions, including the forebrain, retina, midbrain, spinal cord, and the main cell types affected by various diseases in the entire nervous system [10]. For Parkinson's disease, the application of stem cell therapy is under investigation, with the aim of replacing damaged dopamine neurons [11]. However, this approach is still in the research phase.

In conclusion, these therapeutic strategies have varying applications and research stages across different neurodegenerative diseases. While there is currently no definitive cure, these approaches hold promise for alleviating symptoms and slowing disease progression, while enhancing the quality of life for patients. In recent years, numerous researchers have suggested non-pharmacological interventions for treating or preventing various neurodegenerative diseases [12,13,14]. Among them, low-level laser therapy or PBM therapy has attracted much attention due to its beneficial brain effects on animals and humans [15,16,17,18,19,20,21].

3. Photobiomodulation Therapy

PBM is a method that utilizes low-power light from lasers or light-emitting diodes (LEDs) with wavelengths ranging from red to near-infrared (650–1200 nm) to regulate biological function or induce therapeutic effects. Simply put, it is a light-dependent treatment for various diseases. The energy delivered by this treatment method is low enough to prevent any thermal effects. Several previous studies demonstrated the positive impact of PBM on neurodegenerative disorders, including AD, PD, and different types of epilepsy [22,23,24,25]. Despite the successful outcomes of animal studies, few clinical studies that have shown positive outcomes of PBM against neurodegenerative disease have been conducted. Considering the intricate nature of the pathogenesis of neurodegenerative diseases, methods with broad effects, such as PBM, hold promise. Recently, applications of PBM for the nose and brain have shown to be effective in delaying the progress of the neurodegenerative diseases such as AD and PD. In a recent paper, professor Audrey Valverde et al. found that using PBM therapy that applies specific wavelengths of light onto body tissues can have the effect of reducing the neuropathological and behavioral deficits in AD by controlling hypertension [26]. Research has shown that PBM is applied to target organs at specific wavelengths and optimal fluences or energy density to produce therapeutic effects, without any adverse effects

[27,28]. PBM therapy enhances cerebral blood flow, brain energy metabolism, and antioxidant defenses [29,30,31,32]. Additionally, another study group examined the effect of PBM and the application of red to near-infrared light on body tissues, neuroinflammatory responses, and oxidative stress in animal models of neurodegenerative diseases [33]. Neurodegenerative diseases are increasingly prevalent in aging populations worldwide, yet a definitive cure for these conditions remains elusive [34]. Therefore, developing new treatment methods that are safer and more effective in controlling the symptoms of neurodegenerative diseases is an urgent challenge for current research. This review aims to summarize the brain responses to PBM in different preclinical and clinical studies by focusing on mechanisms, neurobiological foundations, and biophysical aspects.

4. Mechanisms of PBM on the Brain

PBM refers to the photochemical and photobiological changes occurring at cellular and molecular levels, mediated by photoreceptors and dependent on the absorption of light by cells [35]. Currently, the light sources relevant to PBM mainly encompass blue light, green light, yellow light, red light, and near-infrared light. Target groups within cells associated with these light sources include cytochrome c oxidase, visual pigments, flavins and flavoproteins, porphyrins, nitrogen-containing compounds, and nitrite

reductases [36].

Cytochrome c oxidase (CCO), a crucial component of the electron transport chain, absorbs photons through interactions with enzymes [37,38]. These interactions cause alterations in proton gradients, triggering an increase in ATP production [39]. They also influence the regulation of reactive oxygen species (ROS) and activation of transcription factors, thereby initiating cell proliferation and inducing cascading signal responses [40,41]. These actions subsequently result in heightened cell proliferation and migration, particularly in fibroblasts. This regulatory process influences cytokines, growth factors, and inflammatory mediators, consequently averting cell apoptosis through the initiation of anti-apoptotic signals. The absorption spectrum of cytochrome c oxidase primarily focuses on red light (630–670 nm) and the near-infrared range (780–940 nm) [42]. Furthermore, beneficial effects of wavelengths such as 1064, 1068, and 1072 nm have been reported in various studies [43,44,45,46]. Given the elevated levels of light scattering within the 800–850 nm range (the primary absorption peak of CCO), it is conceivable that photons may encounter challenges in penetrating deeper tissue layers when compared to longer wavelengths. Nevertheless, within the spectral range of 1064–1072 nm, CCO exhibits reduced light absorption in comparison to the 800–850 nm range. This reduction in absorption, combined with diminished

hemoglobin absorption and reduced light scattering, facilitates the unhindered propagation of light through biological tissues. This phenomenon enables the stimulation of either CCO or ion channels situated within deeper anatomical structures [47,48,49]. Additionally, red light can release various nitric oxide (NO) complexes, including nitrosylmyoglobin, S-nitrosothiols, nitrosylhemoglobin, and nitrosyliron compounds, subsequently affecting mitochondrial function [50].

On the other hand, blue and green light can activate visual pigments, which are G-protein-coupled receptors that can activate transient receptor potential channels [51]. The influx of calcium ions following this activation stimulates calcium-dependent kinase II, phosphorylates the cyclic AMP (cAMP) response element-binding protein in the cell nucleus, and further promotes a series of gene transcription expressions. Flavin mononucleotide exists in complex I of the electron transport chain, where it can perceive blue light to provide activation energy, facilitating the catalysis of oxygen reduction to superoxide [52]. Complex II, another flavin-containing complex, can sense blue light and causes a series of biological effects [53].

5. The Level of Light Penetration into the Brain

As researchers pay more and more attention to PBM

treatment of the brain, some tissue optics laboratories have studied the penetration ability of different wavelengths of light through the scalp and skull as well as the penetration depth of different wavelengths of light in brain parenchyma. This is an interesting issue to consider, as the power density threshold (mW/cm^2) required to cause biological effects on the brain is currently unclear. The ability of light to penetrate biological tissues is closely related to a variety of optical parameters, such as light scattering, absorption, reflection, refraction, wavelength, energy, power, irradiance, radiation exposure (influence), exposure time, continuous wave and pulse wave, irradiation plan, exposure area, distance between the light source and the target tissue, etc. Based on the understanding and analysis of the differences of individual physiology and anatomical structure of tissue samples, as well as the optical properties of various brain regions, it is helpful to select the best optical parameters in PBM treatment to achieve the best penetration effect.

The general principle of light tissue interaction within the 650–1200 nm wavelength optical window most commonly used in PBM is that shorter-wavelength light has lower tissue penetration compared to higher-wavelength light. Ando et al. observed that the penetration rate of an 810 nm laser through the scalp and skull of BALB/c mice is 6.26% [54]. Lapchak et al. compared the transmittance of 810 nm light in four different species of skulls and found that the

transmittance of mice was 40%, while that of rats was 21%, rabbits 11.3%, and human skulls only 4.2% [55]. Salehpour et al. examined light penetration at different wavelengths and found that approximately 23% of the 630 nm laser and 40% of the 810 nm laser reached the surface of the prefrontal cortex of Wistar rats [56]. Zhang et al. observed that a red laser (635 nm) is capable of penetrating through approximately 30% the scalp and skull of C57BL/6 mice [57]. By comparing the penetration ability of light with different wavelengths (660 nm, 808 nm, and 940 nm) in the human cadaveric brain, Tedord et al. found that the penetration ability was the best when using 808 nm wavelength, reaching a depth of 2.52 mm in brain tissue. The brain tissue has a small water absorption peak near the 940 nm wavelength [58]. Beyond the 900 nm wavelength, higher wavelengths lead to higher head penetration, thereby delivering effective levels of energy to deeper brain regions. The comparison of three representative PBM wavelengths, 660 nm, 810 nm, and 1064 nm, for transporting photons to the human head using a Monte Carlo simulation showed that 1064 nm is the optimal wavelength due to its reduced light scattering [59]. Although other photoreceptors (such as water) also absorb photons with higher wavelengths within this range, 1064 nm has less light scattering due to its longer wavelength, thus having deeper penetration ability for the human head. Therefore, in order to achieve the optimal PBM of the human brain, the trade-off between the absorption of

light by CCO and the depth of light penetration must be considered [18].

The debate regarding whether the penetration depth of lasers into tissue is indeed greater than that of LED lights remains unsettled in current scientific discourse. Some researchers have suggested that, in contrast to laser devices generating coherent beams with a narrow focus for penetration, non-coherent LED devices exhibit spatial divergence, limited to superficial layers of tissue [60,61]. Conversely, for LEDs (830 nm) and lasers (810 nm), there appears to be no significant disparity in light penetration through human cranial bones [62]. Further laboratory studies are necessary to elucidate the precise process of light transmission through human skull tissue.

6. Specific Applications of PBM Therapy for Neurodegenerative Diseases

Considering the significant dependence of the brain on mitochondrial activity, it is unsurprising that PBM has undergone extensive testing for the treatment of various brain diseases [63]. PBM activates multiple signaling pathways, including those mediated by ROS, resulting in an upregulation of antioxidant defenses [64]. Anti-apoptotic and pro-survival signals are also triggered [65]. Furthermore, there are two other critical impacts on the ability to shift mitochondrial respiration from glycolysis to

oxidative phosphorylation. Firstly, it mobilizes stem cells from hypoxic niches, allowing them to migrate to sites of injury, where they can facilitate repair. Secondly, alterations in mitochondria can shift the phenotype of macrophages and microglial cells from pro-inflammatory M1 states to anti-inflammatory and phagocytic M2 states [66]. In the brain, the upregulation of neurotrophic factors, such as the brain-derived neurotrophic factor (BDNF), stimulates neurogenesis and fosters synaptic formation and neural plasticity [67,68,69]. Therefore, PBM can regulate several signal transduction and metabolic pathways dependent on neuronal survival. For example, it can improve the survival of neurons, promote neurogenesis, reduce cerebral inflammatory reaction, enhance cerebral blood flow, and maintain mitochondrial homeostasis, so as to improve the response of neurons to cell damage in the process of neurodegenerative disease progression. This review shows that a variety of biological signal transduction and metabolic regulation pathways are affected by PBM in typical neurodegenerative diseases. Combined with the optimal application of specific optical parameters, the molecular response of such cells will be pushed in the right direction ([Figure 1](#)).

[Figure 1](#)

The effects of PBM in neurodegenerative diseases.

6.1. Alzheimer's Disease

6.1.1. Reduction in the Burden of β -Amyloid Plaques

PBM effectively reduces the burden of amyloid plaques by stimulating the phagocytosis of protein aggregates via microglia or by regulating the level of the enzymes involved in the production and degradation pathways of β -amyloid (A β) peptides. PBM promotes microglia phagocytosis by activating Rac1, thus avoiding inflammation-induced cytotoxicity [70]. Professor Wei et al. indicated that the 1070 nm light can induce morphological changes in microglia and increase their colocalization with A β to reduce A β load [71]. A β peptides originate from the sequential proteolytic cleavage of the amyloid precursor protein (APP) by β -secretase, also known as β -site APP cleaving enzyme 1 (BACE1), as well as the γ -secretase complex, comprising presenilin 1 (PS1), nicastrin, and Pen-2. This proteolytic process significantly contributes to the pathological mechanisms underlying Alzheimer's disease [72]. Nonetheless, the generation of A β can be avoided through a nonamyloidogenic pathway involving α -secretase, primarily represented by a disintegrin and metalloproteinase domain-containing protein 10 (ADAM10), in conjunction with the γ -secretase complex [73]. PBM activated the cAMP/PKA/SIRT1 pathway, thereby upregulating ADAM10 and downregulating BACE1, ultimately altering the hydrolysis process of APP.

Additionally, the upregulation of ADAM10 and downregulation of BACE1 by PBM were dependent on SIRT1-coupled retinoic acid receptor β and PPAR γ coactivator 1 α deacetylation [57]. In addition, PBM visibly increased the level of A β -degrading enzyme and insulin degrading enzyme (IDE), but not neutral endopeptidase (NEP), in 5xFAD mice [74]. The study indicated that after internalizing A β ₄₂ in human neuroblastoma cells (SH-EP), irradiation with a 670 nm laser can significantly reduce the accumulation of A β ₄₂ in the cells [75].

6.1.2. Neuroinflammation

Glial cells represent a substantial category of cells in the nervous tissue distinct from neurons. They are broadly distributed in both the central and peripheral nervous systems and primarily encompass astrocytes, oligodendrocytes (collectively referred to as macroglia), and microglia [76]. Microglial cells display two distinct phenotypes in response to inflammation. In the absence of pathogens, microglia remain quiescent. However, upon the detection of pathological hallmarks of AD, microglia become activated, releasing inflammatory mediators, and participate in the clearance of A β plaques. Activated microglia manifest in either a pro-inflammatory or an anti-inflammatory state [77]. M1 microglia assume a pro-inflammatory state, continuously releasing inflammatory factors such as ROS

and NO, which, through excessive pruning of neuronal synapses and demyelination, ultimately result in disrupted information transmission and even cell death [78]. In contrast, M2 microglia adopt an anti-inflammatory state, capable of phagocytosing A β plaques, providing trophic factors, promoting neural network reconstruction, and facilitating brain repair in response to injury [79,80].

In various neurodegenerative diseases, including AD, PBM plays a crucial role in modulating central nervous system damage mediated by microglial cells. Irradiating primary microglial cells and microglia-like BV-2 cells with an 808 nm LED at different doses, it was observed that at higher doses (4–30 J/cm²), microglial cells predominantly transitioned to the M1 phenotype, whereas at lower doses (0.2–10 J/cm²), they primarily shifted towards the M2 phenotype [77]. Using a 633 nm laser, Song et al. [70] explored the effect of PBM on the neurotoxicity induced by lipopolysaccharide (LPS)-activated microglial cells. The results revealed that PBM can inhibit the activation of LPS-stimulated microglial cells by activating the non-receptor tyrosine kinase Src. This inhibition reduced the neurotoxic effects caused by microglial cells, enhanced their phagocytic capabilities, and consequently facilitated the clearance of A β aggregates. Furthermore, Professor Qu's research group utilized 808 nm laser irradiation on primary microglial cells from the mouse hippocampus, which were activated by A β_{1-42} oligomers

[81]. The results indicated that in M1-type activated microglial cells induced by A β oligomers, PBM effectively induced a metabolic shift from glycolysis to mitochondrial activity. This shift led to the reduced secretion of pro-inflammatory factors and enhanced phagocytic function, transitioning the microglial cells towards the M2 phenotype. Simultaneously, in a co-culture model of neurons and activated microglial cells, PBM was found to effectively reduce neuronal apoptosis by modulating the levels of ROS generated by A β -activated microglial cells.

Furthermore, similar to microglial cells, astrocytes also exhibit two states, pro-inflammatory (A1) and anti-inflammatory (A2), during CNS inflammation. A1 astrocytes release pro-inflammatory factors such as NO and ROS, which exert toxicity in glial cells, rendering them phagocytosis-impaired and incapable of clearing A β plaques [82]. A2 astrocytes release ATP, γ -aminobutyric acid (GABA), neurotrophic factors, and cytokines, promoting neuronal proliferation, differentiation, and synaptic repair [83]. Early studies revealed that 632.8 nm laser pretreatment can suppress superoxide dismutase activity in A β_{1-42} -induced astrocytes, reducing the expression of interleukin-1 β (IL-1 β) and inducible nitric oxide synthase (iNOS), thereby exerting an anti-neuroinflammatory effect [84]. Silveira et al. [85] found that irradiating C6 astroglioma cells with a 660 nm laser led to increased respiratory chain

complex I activity and hexokinase activity, resulting in alterations in cytoplasmic and mitochondrial redox states, enhanced ATP production, and significantly improved C6 cell mitochondrial activity. Yoon et al. [86] discovered that treating primary astrocytes from SD rats with a 660 nm LED increased the co-expression of GFAP and bromodeoxyuridine (BrdU)/Ki67, promoting specific proliferation. Additionally, the co-expression of Oct4 and GFAP decreased, while Nestin and ALDH1L1 expression increased, suggesting that a 660 nm LED can facilitate astrocyte differentiation. These findings indicate that PBM can modify astrocyte activity, promoting their proliferation or differentiation, thereby ameliorating the pathological state of the CNS.

For the in vivo test, Blivet et al. [57] applied a transcranial pulse PBM using a combination of red and NIR light, causing a reduction in hippocampal inflammation markers TNF- α , IL-6, and IL-1 β . Simultaneously, it decreased the activation of astrocytes and microglial cells, leading to the restoration of spatial working memory in the Y-maze test for mice.

Professor Wei Xunbin et al. used continuous LED devices emitting light in the range from 1040 nm to 1090 nm for transcranial PBM in APP/PS1 mice. This treatment significantly improved spatial learning and memory in the Morris water maze test for AD mice and reduced the area of A β plaques in the hippocampal region [87]. In 2021, the

same team conducted transcranial PBM on AD model mice using 10 Hz and 40 Hz 1070 nm near-infrared light. The results indicate that both 10 Hz and 40 Hz 1070 nm near-infrared light can reduce the production of M1-like microglial cells, alleviating the inflammatory response in AD model mice [71]. In 2016, the results of Li-Huei Tsai et al. showed that a 40 Hz LED (white light) induced gamma oscillations in the visual cortex of the brain, leading to morphological changes in microglial cells in AD mice and subsequently enhancing amyloid protein phagocytosis to reduce the occurrence of AD [88]. In 2019, their team explored the effectiveness of combined visual and auditory stimulation at 40 Hz. They discovered that this combined stimulation could promote the clustering of microglial cells around A β plaques, leading to a widespread reduction in amyloid proteins and phosphorylated Tau proteins in multiple brain regions [89]. This, in turn, improved spatial memory and cognitive abilities in AD mice. In the same year, the research group investigated the mechanism of long-term 40 Hz light therapy. The results reveal that long-term 40 Hz light therapy induced gamma oscillations in multiple regions of the brain, including the visual cortex, hippocampus, and frontal cortex. This led to changes in the expression of genes related to neurons, synaptic damage, and microglial cell morphology, mitigating neuron damage and microglial cell inflammatory responses, improving synaptic function, enhancing neuroprotection, and ultimately enhancing

cognitive function in AD mice [90]. Furthermore, PBM promoted the level of IL-3Ra in microglial cells and IL-3 in astrocytes, facilitating the recruitment of microglial cells around amyloid plaques [91].

6.1.3. Neurogenesis

The CNS comprises neural stem cells (NSCs), which produce various neurons and glia and seem to be very suitable as alternative therapies for various cell loss diseases [92]. Under the pathological conditions of neurodegenerative diseases, especially during the development of AD, adult hippocampal neurogenesis (AHN) is inhibited, which reduces the stability of the neural circuit and leads to a decrease in the production of new neurons in the dentate gyrus (DG), making it difficult for AD to recover from learning/memory dysfunction. Professor Da Xing's research group found that PBM facilitated adult hippocampal neurogenesis in AD mice. Both *in vitro* and *in vivo*, PBM triggered the activation of latent transforming growth factor- β 1 (LTGF β 1) during AHN induction. This activation subsequently enhanced the differentiation of NSCs in the hippocampus of APP/PS1 mice, promoting their transition into newly generated neurons [93]. Numerous findings have shown that the immune system is closely related to the synaptic plasticity, inflammation, and progression of AD [94,95,96,97,98,99]. However, few studies have improved

the niche microenvironment of NSCs in the brain of AD patients by regulating the function of nonparenchymal immune cells, in order to promote AHN. Therefore, in 2022, for the first time, their research team found that PBM regulates peripheral CD4⁺ T lymphocytes, which contributes to promoting adult hippocampal neurogenesis and alleviating AD cognitive impairment [100]. Furthermore, a descriptive report on dementia care from Ivan V Maksimovich et al. showed that 48 patients suffering from AD received transcatheter intracerebral laser PBM through catheterization to stimulate neurogenesis and tissue structure recovery, improving cerebral blood flow and leading to a reduction in dementia and enhancement of cognitive function, an effect that can last for a long time [101].

6.1.4. Apoptosis

Cell apoptosis stands as one of the pathophysiological mechanisms in normal brain aging and neurodegenerative diseases. The phenomenon of PBM anti-apoptotic properties was initially observed in skeletal muscle satellite cells by Shefer et al. [102]. The study highlighted that laser irradiation at a wavelength of 632.8 nm could diminish the levels of p53, p21, and Bax, while concurrently elevating Bcl-2 levels 24 h post-irradiation, thus safeguarding skeletal muscle satellite cells from apoptosis. Additionally, LED light

at 640 nm inhibited apoptosis in PC12 cells induced by A β toxicity, as evidenced 24 h post-irradiation [103]. Light at 670 nm (LED) [104] and 810 nm (laser) [105] substantially prevented apoptosis by decreasing pro-apoptotic factors as well as by inhibiting Caspase-3. One plausible explanation for the neuroprotective effect of this light could be attributed to the initial mechanism of light absorption by chromophores within the mitochondrial inner membrane enzymes, leading to an enhancement in mitochondrial membrane potential (MMP) [106]. Furthermore, Zhang and colleagues demonstrated that low-dose laser irradiation at 632.8 nm effectively reversed apoptosis in PC12 cells by reducing the mRNA Bax/Bcl-xl ratio through the activation of the PKC pathway [107]. Laser irradiation at 632.8 nm notably suppressed the activation of glycogen synthase kinase 3 β , Bax, and Caspase-3, effectively averting staurosporine-induced apoptosis [108].

6.1.5. Cerebral Blood Flow and Neurotrophic Factors

Nitric oxide (NO) is a potent vasodilator that may be released from its binding sites through a photodissociation process within the respiratory chain when undergoing PBM. Based on preclinical research, PBM has the capacity to elevate the levels of neuronal NO, enlarge blood vessel diameter, and enhance cerebral blood flow (CBF) [109,110]. Consequently, it is plausible to consider that PBM therapy targeted at

specific brain regions may have the potential to influence regional CBF [111]. Transcranial PBM utilizing red and NIR wavelengths resulted in an elevation of cerebral oxygen consumption in both naive rats and A β PP mice [112]. Additionally, in the latest clinical studies conducted by Hanli Liu et al., enhancements in cerebral oxygenation and hemodynamics were noted during and after transcranial laser irradiation at 1064 nm [30,47].

Among the various members within the neurotrophic factors family, known as neurotrophins, research has shifted towards exploring the stimulating impacts of PBM on BDNF, nerve growth factor (NGF), and glial cell-derived neurotrophic factor (GDNF). The upregulation of neurotrophins such as BDNF and NGF could explain the observed enhancements in neurogenesis and synaptogenesis [113]. Increasing BDNF expression may contribute to the attenuation of cortical dendrite atrophy in the CNS during the progression of AD [69]. In this context, it has been proposed that PBM (632.8 nm) can rescue dendritic atrophy by activating the ERK/CREB/BDNF pathway [69]. In similar research employing the same laser wavelength (632.8 nm), PBM triggered IP3 receptor activation, leading to elevated Ca²⁺ levels. Subsequently, this activation promoted the ERK/CREB pathway, ultimately culminating in heightened expression of BDNF [114]. For in vivo experiments, a coherent laser light at 670 nm

significantly elevated BDNF in the cortex of rats [104].

Currently, despite the substantial financial investments made in the development of therapeutic drugs for AD, significant breakthroughs in its treatment efficacy remain elusive. However, the therapeutic efficacy and safety of PBM have been validated in numerous preclinical and clinical studies ([Table 1](#)), suggesting that PBM may emerge as a promising alternative therapeutic approach for neurodegenerative diseases. By leveraging advantages such as a non-invasive nature and lack of side effects, PBM can serve as an adjunctive therapy to existing effective treatments and even evolve into a preventive therapeutic modality. In summary, PBM holds great promise in the treatment and prevention of cognitive impairments.

Table 1

Summary of in vivo studies on the effects of photobiomodulation therapy in Alzheimer's disease.

Disease	Index	Irradiation Parameters		
	Light Source	Wavelength	Continuous/Frequency	

	LED	473 nm	CW; 40 Hz
	LED	500 nm	CW
Alzheimer's disease	LED	610–670 nm	CW; 40 Hz
	Fiber laser	808/810 nm	CW
	Fiber laser	808/810 nm	10–100 Hz
	Fiber laser, LED	1040–1267 nm	CW; 10, 40, 600 Hz

6.2. Parkinson's Disease

Parkinson's disease is a multifactorial, multisystem disorder characterized by the loss of dopaminergic neurons in the substantia nigra pars compacta (SNc) of the brain [137]. The depletion of dopamine (DA) predominantly influences motor

function, giving rise to symptoms such as bradykinesia, muscle rigidity, resting tremors, and postural instability. However, it also manifests in various other signs and symptoms affecting emotions, cognition, the digestive system, and the sense of smell. The death of dopaminergic neurons is attributed to the accumulation of Lewy bodies, which are aggregates of α -synuclein, within the cells, although this theory remains somewhat controversial. The exact cause of PD is not entirely understood. A genetic component is observed in only around 15% of PD patients, with the most common mutations occurring in genes such as leucine-rich repeat kinase 2, glucocerebrosidase, and α -synuclein [138,139].

Numerous studies have presented promising outcomes regarding the application of PBM in animal models of Parkinson's disease (refer to [Table 2](#)). A recent review of animal evidence has indicated that human trials are warranted based on the encouraging findings [140]. Photobiomodulation has demonstrated efficacy in pre-treating and safeguarding animals, including non-human primates, from the effects of toxin (MPTP)-induced Parkinson's disease models. This protection extends to mitigating induced PD signs and preserving substantia nigra neurons [141,142]. Previous studies show that PBM restored DA levels in toxin (MPP+)-exposed cells by increasing the expression of the dopamine transport protein VMAT2 and

tyrosine hydroxylase, the enzyme responsible for DA synthesis. The regulation of VMAT2 expression occurred via the ERK/CREB pathway, a critical regulator in maintaining, releasing, and expressing monoamines like dopamine and serotonin in the central nervous system monoaminergic neurons, thereby underscoring the neuroprotective effects of PBM [143,144,145]. Changes in mitochondrial dynamics, such as mitochondrial fission, fragmentation, and functional loss, are commonly observed in PD [146]. Post-treatment with PBM preserved healthy mitochondrial dynamics and suppressed the mitochondrial fragmentation in CA1 neurons. This treatment reduced the detrimental activity of dynamin-related protein 1 (Drp1) GTPase, mitochondrial fission factor (Mff), and mitochondrial fission 1 protein (Fis1) and balanced mitochondrial targeting fission and fusion protein in global cerebral ischemia [147]. Moreover, there are reports indicating that twice-daily LED treatment (at 670 nm) markedly reduced the count of striatal and cortical neurons undergoing apoptosis induced by exposure to rotenone and MPP+ [148]. Findings from a mouse model of PD reveal that the application of photobiomodulation at 670 nm to distal tissues (excluding the head) exerts a notable *ex vivo* neuroprotective effect by mitigating the loss of midbrain dopaminergic neurons [149]. The pathogenesis of PD is associated with abnormalities in the SNc, a midbrain structure located at an approximate depth of 80–100 mm from the coronal suture, beneath the dura mater. Research

suggests that near-infrared light may not penetrate the human brain effectively, with a depth of penetration not exceeding 20 mm beyond the cortical surface [150]. This limitation is deemed noteworthy in the context of transcranial photobiomodulation therapy for Parkinson's disease.

Table 2

Summary of *in vivo* studies on the effects of photobiomodulation therapy in Parkinson's disease.

	Index	Irradiation Param		
Disease		Light Source	Wavelength	Continuous/F
Parkinson's disease		Laser	405 nm	CW
		Fiber laser, LED	627–675 nm	CW
		Semiconductor laser, LED	808–810 nm	CW
		Semiconductor laser	904 nm	50 H

7. Conclusions and Future Perspectives

A substantial body of research on PBM has been published, including investigations into its mechanisms of action and methods for assessing therapeutic outcomes, providing a strong evidence-based foundation for future clinical trials. However, PBM has a long journey ahead before becoming a widely accessible therapeutic approach. Over the past decade, PBM has been extensively employed in drug development for various diseases, such as AD, demonstrating favorable therapeutic effects. Nevertheless, several unresolved issues have been identified. Currently, its mechanisms of action and dose-response relationships, especially the dose-response relationship in AD treatment, remain unclear. More comprehensive and in-depth clinical trials are required to elucidate the optimal treatment modalities and dosage parameters to adopt in clinical practice and to refine the clinical assessment system. Simultaneously, there is a need to develop portable PBM treatment devices tailored for neurodegenerative diseases, offering a promising new direction for clinical treatment and a feasible solution to alleviate the burdens of disease management and healthcare costs for patients.

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Author Contributions

Q.S. conceived the article and guided the study; H.G. and Y.Y. performed the literature search and analysis, wrote the paper, and designed the figures and tables. The final draft was critically revised by Q.S. All authors have read and agreed to the published version of the manuscript.

Conflicts of Interest

The authors declare no conflicts of interest.

Footnotes

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References

1. Lynch C. World Alzheimer report 2019: Attitudes to dementia, a global survey. *Alzheimers Dement.* 2020;16:e038255. doi: 10.1002/alz.038255. [[CrossRef](#)] [[Google Scholar](#)]
2. Dorsey E.R., Bloem B.R. The Parkinson pandemic—A call to action. *JAMA Neurol.* 2018;75:9–10. doi: 10.1001/jamaneurol.2017.3299. [[PubMed](#)] [[CrossRef](#)] [[Google Scholar](#)]
3. Hardiman O., Al-Chalabi A., Chio A., Corr E.M., Logroscino G., Robberecht W., Shaw P.J., Simmons Z., van den Berg L.H. Amyotrophic lateral sclerosis. *Nat. Rev. Dis. Primers.* 2017;3:17071. doi: 10.1038/nrdp.2017.71. [[PubMed](#)] [[CrossRef](#)] [[Google Scholar](#)]
4. Bellou V., Belbasis L., Tzoulaki I., Evangelou E., Ioannidis J.P.A. Environmental risk factors and Parkinson's disease: An umbrella review of meta-analyses. *Park. Relat. Disord.* 2016;23:1–9. doi: 10.1016/j.parkreldis.2015.12.008. [[PubMed](#)] [[CrossRef](#)] [[Google Scholar](#)]
5. Cummings J., Lee G., Ritter A., Sabbagh M., Zhong K. Alzheimer's disease drug development pipeline: 2020. *Alzheimer's Dement. Transl. Res. Clin. Interv.* 2020;6:e12050. doi: 10.1002/trc2.12050. [[PMC free article](#)] [[PubMed](#)] [[CrossRef](#)] [[Google Scholar](#)]
6. Connolly B.S., Lang A.E. Pharmacological treatment of Parkinson disease: A review. *JAMA.* 2014;311:1670–1683.

doi: 10.1001/jama.2014.3654. [PubMed] [CrossRef] [Google Scholar]

7. Sun J., Roy S. Gene-based therapies for neurodegenerative diseases. *Nat. Neurosci.* 2021;24:297–311. doi: 10.1038/s41593-020-00778-1. [PMC free article] [PubMed] [CrossRef] [Google Scholar]
8. Lee J.-M., Correia K., Loupe J., Kim K.-H., Barker D., Hong E.P., Chao M.J., Long J.D., Lucente D., Vonsattel J.P.G., et al. CAG repeat not polyglutamine length determines timing of Huntington's disease onset. *Cell.* 2019;178:887–900.e814. doi: 10.1016/j.cell.2019.06.036. [PMC free article] [PubMed] [CrossRef] [Google Scholar]
9. Song C., Shi J., Zhang P., Zhang Y., Xu J., Zhao L., Zhang R., Wang H., Chen H. Immunotherapy for Alzheimer's disease: Targeting β-amyloid and beyond. *Transl. Neurodegener.* 2022;11:18. doi: 10.1186/s40035-022-00292-3. [PMC free article] [PubMed] [CrossRef] [Google Scholar]
10. Temple S. Advancing cell therapy for neurodegenerative diseases. *Cell Stem Cell.* 2023;30:512–529. doi: 10.1016/j.stem.2023.03.017. [PMC free article] [PubMed] [CrossRef] [Google Scholar]
11. Barker R.A., Farrell K., Guzman N.V., He X., Lazic S.E., Moore S., Morris R., Tyers P., Wijeyekoon R., Daft D., et al. Designing stem-cell-based dopamine cell replacement trials for Parkinson's disease. *Nat. Med.* 2019;25:1045–1053. doi: 10.1038/s41591-019-0507-2. [PubMed] [CrossRef]

[\[Google Scholar\]](#)

12. Gonzalez-Lima F., Barksdale B.R., Rojas J.C. Mitochondrial respiration as a target for neuroprotection and cognitive enhancement. *Biochem. Pharmacol.* 2014;88:584–593. doi: 10.1016/j.bcp.2013.11.010. [\[PubMed\]](#) [\[CrossRef\]](#)

[\[Google Scholar\]](#)

13. Salgado A.S.I., Zângaro R.A., Parreira R.B., Kerppers I.I. The effects of transcranial LED therapy (TCLT) on cerebral blood flow in the elderly women. *Lasers Med. Sci.* 2015;30:339–346. doi: 10.1007/s10103-014-1669-2.

[\[PubMed\]](#) [\[CrossRef\]](#) [\[Google Scholar\]](#)

14. De la Torre J.C., Olmo A.D., Valles S. Can mild cognitive impairment be stabilized by showering brain mitochondria with laser photons? *Neuropharmacology*. 2020;171:107841. doi: 10.1016/j.neuropharm.2019.107841. [\[PubMed\]](#)

[\[CrossRef\]](#) [\[Google Scholar\]](#)

15. Rojas J.C., Gonzalez-Lima F. Neurological and psychological applications of transcranial lasers and LEDs. *Biochem. Pharmacol.* 2013;86:447–457.

doi: 10.1016/j.bcp.2013.06.012. [\[PubMed\]](#) [\[CrossRef\]](#)

[\[Google Scholar\]](#)

16. Salehpour F., Ahmadian N., Rasta S.H., Farhoudi M., Karimi P., Sadigh-Eteghad S. Transcranial low-level laser therapy improves brain mitochondrial function and cognitive impairment in D-galactose–induced aging mice. *Neurobiol. Aging*. 2017;58:140–150.

doi: 10.1016/j.neurobiolaging.2017.06.025. [\[PubMed\]](#)

[[CrossRef](#)] [[Google Scholar](#)]

17. Salehpour F., De Taboada L., Cassano P., Kamari F., Mahmoudi J., Ahmadi-Kandjani S., Rasta S.H., Sadigh-Eteghad S. A protocol for transcranial photobiomodulation therapy in mice. *JoVE*. 2018:e59076. [[PubMed](#)] [[Google Scholar](#)]
18. Saucedo C.L., Courtois E.C., Wade Z.S., Kelley M.N., Kheradbin N., Barrett D.W., Gonzalez-Lima F. Transcranial laser stimulation: Mitochondrial and cerebrovascular effects in younger and older healthy adults. *Brain Stimul.* 2021;14:440–449. doi: 10.1016/j.brs.2021.02.011. [[PubMed](#)] [[CrossRef](#)] [[Google Scholar](#)]
19. Vargas E., Barrett D.W., Saucedo C.L., Huang L.-D., Abraham J.A., Tanaka H., Haley A.P., Gonzalez-Lima F. Beneficial neurocognitive effects of transcranial laser in older adults. *Lasers Med. Sci.* 2017;32:1153–1162. doi: 10.1007/s10103-017-2221-y. [[PMC free article](#)] [[PubMed](#)] [[CrossRef](#)] [[Google Scholar](#)]
20. dos Santos Cardoso F., dos Santos J.C.C., Gonzalez-Lima F., Araújo B.H.S., Lopes-Martins R.Á.B., Gomes da Silva S. Effects of chronic photobiomodulation with transcranial near-infrared laser on brain metabolomics of young and aged rats. *Mol. Neurobiol.* 2021;58:2256–2268. doi: 10.1007/s12035-020-02247-z. [[PubMed](#)] [[CrossRef](#)] [[Google Scholar](#)]
21. Cardoso F.d.S., de Souza Oliveira Tavares C., Araujo B.H.S., Mansur F., Lopes-Martins R.Á.B., Gomes da Silva S.

- Improved spatial memory and neuroinflammatory profile changes in aged rats submitted to photobiomodulation therapy. *Cell Mol. Neurobiol.* 2022;42:1875–1886.
doi: 10.1007/s10571-021-01069-4. [PubMed] [CrossRef] [Google Scholar]
22. Hong N. Photobiomodulation as a treatment for neurodegenerative disorders: Current and future trends. *Biomed. Eng. Lett.* 2019;9:359–366. doi: 10.1007/s13534-019-00115-x. [PMC free article] [PubMed] [CrossRef] [Google Scholar]
23. Liebert A., Bicknell B., Laakso E.L., Heller G., Jalilitabaei P., Tilley S., Mitrofanis J., Kiat H. Improvements in clinical signs of Parkinson's disease using photobiomodulation: A prospective proof-of-concept study. *BMC Neurol.* 2021;21:256. doi: 10.1186/s12883-021-02248-y. [PMC free article] [PubMed] [CrossRef] [Google Scholar]
24. Su M., Nizamutdinov D., Liu H., Huang J.H. Recent mechanisms of neurodegeneration and photobiomodulation in the context of Alzheimer's disease. *Int. J. Mol. Sci.* 2023;24:9272. doi: 10.3390/ijms24119272. [PMC free article] [PubMed] [CrossRef] [Google Scholar]
25. Bicknell B., Liebert A., Borody T., Herkes G., McLachlan C., Kiat H. Neurodegenerative and neurodevelopmental diseases and the gut-brain axis: The potential of therapeutic targeting of the microbiome. *Int. J. Mol. Sci.* 2023;24:9577. doi: 10.3390/ijms24119577. [PMC free article] [PubMed] [CrossRef] [Google Scholar]

26. Valverde A., Mitrofanis J. Photobiomodulation for hypertension and Alzheimer's disease. *J. Alzheimers Dis.* 2022;90:1045–1055. doi: 10.3233/JAD-220632. [[PMC free article](#)] [[PubMed](#)] [[CrossRef](#)] [[Google Scholar](#)]
27. Santana-Blank L., Rodríguez-Santana E., Santana-Rodríguez K.E., Reyes H. “Quantum Leap” in photobiomodulation therapy ushers in a new generation of light-based treatments for cancer and other complex diseases: Perspective and mini-review. *Photomed. Laser Surg.* 2016;34:93–101. doi: 10.1089/pho.2015.4015. [[PMC free article](#)] [[PubMed](#)] [[CrossRef](#)] [[Google Scholar](#)]
28. Chen Y., De Taboada L., O'Connor M., Delapp S., Zivin J.A. Thermal effects of transcranial near-infrared laser irradiation on rabbit cortex. *Neurosci. Lett.* 2013;553:99–103. doi: 10.1016/j.neulet.2013.07.049. [[PubMed](#)] [[CrossRef](#)] [[Google Scholar](#)]
29. Rojas J.C., Bruchey A.K., Gonzalez-Lima F. Low-level light therapy improves cortical metabolic capacity and memory retention. *J. Alzheimers Dis.* 2012;32:741–752. doi: 10.3233/JAD-2012-120817. [[PubMed](#)] [[CrossRef](#)] [[Google Scholar](#)]
30. Tian F., Hase S.N., Gonzalez-Lima F., Liu H. Transcranial laser stimulation improves human cerebral oxygenation. *Lasers Surg. Med.* 2016;48:343–349. doi: 10.1002/lsm.22471. [[PMC free article](#)] [[PubMed](#)] [[CrossRef](#)] [[Google Scholar](#)]
31. Nawashiro H., Wada K., Nakai K., Sato S. Focal increase

in cerebral blood flow after treatment with near-infrared light to the forehead in a patient in a persistent vegetative state.

Photomed. Laser Surg. 2011;30:231–233.

doi: 10.1089/pho.2011.3044. [[PubMed](#)] [[CrossRef](#)] [[Google Scholar](#)]

32. Lu Y., Wang R., Dong Y., Tucker D., Zhao N., Ahmed M.E., Zhu L., Liu T.C.-Y., Cohen R.M., Zhang Q. Low-level laser therapy for beta amyloid toxicity in rat hippocampus.

Neurobiol. Aging. 2017;49:165–182.

doi: 10.1016/j.neurobiolaging.2016.10.003. [[PMC free article](#)] [[PubMed](#)] [[CrossRef](#)] [[Google Scholar](#)]

33. Xie K., El Khoury H., Mitrofanis J., Austin P.J. A systematic review of the effect of photobiomodulation on the neuroinflammatory response in animal models of neurodegenerative diseases. *Rev. Neurosci.* 2023;34:459–481. doi: 10.1515/revneuro-2022-0109. [[PubMed](#)] [[CrossRef](#)] [[Google Scholar](#)]

34. Abijo A., Lee C.-Y., Huang C.-Y., Ho P.-C., Tsai K.-J. The beneficial role of photobiomodulation in neurodegenerative diseases. *Biomedicines.* 2023;11:1828.

doi: 10.3390/biomedicines11071828. [[PMC free article](#)] [[PubMed](#)] [[CrossRef](#)] [[Google Scholar](#)]

35. Cardoso F.d.S., Gonzalez-Lima F., Gomes da Silva S. Photobiomodulation for the aging brain. *Ageing Res. Rev.* 2021;70:101415. doi: 10.1016/j.arr.2021.101415. [[PubMed](#)] [[CrossRef](#)] [[Google Scholar](#)]

36. Salehpour F., Mahmoudi J., Kamari F., Sadigh-Eteghad

- S., Rasta S.H., Hamblin M.R. Brain photobiomodulation therapy: A narrative review. *Mol. Neurobiol.* 2018;55:6601–6636. doi: 10.1007/s12035-017-0852-4. [[PMC free article](#)] [[PubMed](#)] [[CrossRef](#)] [[Google Scholar](#)]
37. Karu T. Primary and secondary mechanisms of action of visible to near-IR radiation on cells. *J. Photochem. Photobiol. B Biol.* 1999;49:1–17. doi: 10.1016/S1011-1344(98)00219-X. [[PubMed](#)] [[CrossRef](#)] [[Google Scholar](#)]
38. Karu T. *Biomedical Photonics Handbook*. Vol. 48 CRC Press; Boca Raton, FL, USA: 2003. Low-Power Laser Therapy. [[Google Scholar](#)]
39. Wong-Riley M.T.T., Liang H.L., Eells J.T., Chance B., Henry M.M., Buchmann E., Kane M., Whelan H.T. Photobiomodulation directly benefits primary neurons functionally inactivated by toxins: Role of cytochrome c oxidase. *J. Biol. Chem.* 2005;280:4761–4771. doi: 10.1074/jbc.M409650200. [[PubMed](#)] [[CrossRef](#)] [[Google Scholar](#)]
40. Suski J., Lebiedzinska M., Bonora M., Pinton P., Duszynski J., Wieckowski M.R. *Mitochondrial Bioenergetics*. Springer; Berlin/Heidelberg, Germany: 2018. Relation between mitochondrial membrane potential and ROS formation; pp. 357–381. [[PubMed](#)] [[Google Scholar](#)]
41. Waypa G.B., Smith K.A., Schumacker P.T. O₂ sensing, mitochondria and ROS signaling: The fog is lifting. *Mol. Asp. Med.* 2016;47–48:76–89. doi: 10.1016/j.mam.2016.01.002. [[PMC free article](#)] [[PubMed](#)] [[CrossRef](#)] [[Google Scholar](#)]

42. Karu T.I., Kolyakov S.F. Exact action spectra for cellular responses relevant to phototherapy. *Photomed. Laser Surg.* 2005;23:355–361. doi: 10.1089/pho.2005.23.355. [[PubMed](#)] [[CrossRef](#)] [[Google Scholar](#)]
43. Barrett D.W., Gonzalez-Lima F. Transcranial infrared laser stimulation produces beneficial cognitive and emotional effects in humans. *Neuroscience*. 2013;230:13–23. doi: 10.1016/j.neuroscience.2012.11.016. [[PubMed](#)] [[CrossRef](#)] [[Google Scholar](#)]
44. Blanco N.J., Maddox W.T., Gonzalez-Lima F. Improving executive function using transcranial infrared laser stimulation. *J. Neuropsychol.* 2017;11:14–25. doi: 10.1111/jnp.12074. [[PMC free article](#)] [[PubMed](#)] [[CrossRef](#)] [[Google Scholar](#)]
45. Hwang J., Castelli D.M., Gonzalez-Lima F. Cognitive enhancement by transcranial laser stimulation and acute aerobic exercise. *Lasers Med. Sci.* 2016;31:1151–1160. doi: 10.1007/s10103-016-1962-3. [[PubMed](#)] [[CrossRef](#)] [[Google Scholar](#)]
46. Dougal G., Lee S.Y. Evaluation of the efficacy of low-level light therapy using 1072 nm infrared light for the treatment of herpes simplex labialis. *Clin. Exp. Dermatol.* 2013;38:713–718. doi: 10.1111/ced.12069. [[PubMed](#)] [[CrossRef](#)] [[Google Scholar](#)]
47. Wang X., Tian F., Reddy D.D., Nalawade S.S., Barrett D.W., Gonzalez-Lima F., Liu H. Up-regulation of cerebral cytochrome-c-oxidase and hemodynamics by transcranial

- infrared laser stimulation: A broadband near-infrared spectroscopy study. *J. Cereb. Blood Flow. Metab.* 2017;37:3789–3802. doi: 10.1177/0271678X17691783. [[PMC free article](#)] [[PubMed](#)] [[CrossRef](#)] [[Google Scholar](#)]
48. Wang X., Tian F., Soni S.S., Gonzalez-Lima F., Liu H. Interplay between up-regulation of cytochrome-c-oxidase and hemoglobin oxygenation induced by near-infrared laser. *Sci. Rep.* 2016;6:30540. doi: 10.1038/srep30540. [[PMC free article](#)] [[PubMed](#)] [[CrossRef](#)] [[Google Scholar](#)]
49. Bradford A., Barlow A., Chazot P.L. Probing the differential effects of infrared light sources IR1072 and IR880 on human lymphocytes: Evidence of selective cytoprotection by IR1072. *J. Photochem. Photobiol. B Biol.* 2005;81:9–14. doi: 10.1016/j.jphotobiol.2005.05.005. [[PubMed](#)] [[CrossRef](#)] [[Google Scholar](#)]
50. Karu T.I., Pyatibrat L.V., Afanasyeva N.I. Cellular effects of low power laser therapy can be mediated by nitric oxide. *Lasers Surg. Med.* 2005;36:307–314. doi: 10.1002/lsm.20148. [[PubMed](#)] [[CrossRef](#)] [[Google Scholar](#)]
51. Kaylor J.J., Xu T., Ingram N.T., Tsan A., Hakobyan H., Fain G.L., Travis G.H. Blue light regenerates functional visual pigments in mammals through a retinyl-phospholipid intermediate. *Nat. Commun.* 2017;8:16. doi: 10.1038/s41467-017-00018-4. [[PMC free article](#)] [[PubMed](#)] [[CrossRef](#)] [[Google Scholar](#)]
52. Lukacs A., Tonge P.J., Meech S.R. Photophysics of the

- blue light using flavin domain. *Acc. Chem. Res.* 2022;55:402–414. doi: 10.1021/acs.accounts.1c00659. [\[PubMed\]](#) [\[CrossRef\]](#) [\[Google Scholar\]](#)
53. Yang M.-Y., Chang C.-J., Chen L.-Y. Blue light induced reactive oxygen species from flavin mononucleotide and flavin adenine dinucleotide on lethality of HeLa cells. *J. Photochem. Photobiol. B Biol.* 2017;173:325–332. doi: 10.1016/j.jphotobiol.2017.06.014. [\[PubMed\]](#) [\[CrossRef\]](#) [\[Google Scholar\]](#)
54. Ando T., Xuan W., Xu T., Dai T., Sharma S.K., Kharkwal G.B., Huang Y.-Y., Wu Q., Whalen M.J., Sato S., et al. Comparison of therapeutic effects between pulsed and continuous wave 810-nm wavelength laser irradiation for traumatic brain injury in mice. *PLoS ONE*. 2011;6:e26212. doi: 10.1371/journal.pone.0026212. [\[PMC free article\]](#) [\[PubMed\]](#) [\[CrossRef\]](#) [\[Google Scholar\]](#)
55. Lapchak P.A., Boitano P.D., Butte P.V., Fisher D.J., Hölscher T., Ley E.J., Nuño M., Voie A.H., Rajput P.S. Transcranial near-infrared laser transmission (NILT) profiles (800 nm): Systematic comparison in four common research species. *PLoS ONE*. 2015;10:e0127580. doi: 10.1371/journal.pone.0127580. [\[PMC free article\]](#) [\[PubMed\]](#) [\[CrossRef\]](#) [\[Google Scholar\]](#)
56. Salehpour F., Rasta S.H., Mohaddes G., Sadigh-Eteghad S., Salarirad S. Therapeutic effects of 10-Hz pulsed wave lasers in rat depression model: A comparison between near-infrared and red wavelengths. *Lasers Surg. Med.*

2016;48:695–705. doi: 10.1002/lsm.22542. [[PubMed](#)] [[CrossRef](#)] [[Google Scholar](#)]

57. Zhang Z., Shen Q., Wu X., Zhang D., Xing D. Activation of PKA/SIRT1 signaling pathway by photobiomodulation therapy reduces A β levels in Alzheimer's disease models. *Aging Cell.* 2020;19:e13054. doi: 10.1111/acel.13054. [[PMC free article](#)] [[PubMed](#)] [[CrossRef](#)] [[Google Scholar](#)]

58. Tedford C.E., DeLapp S., Jacques S., Anders J. Quantitative analysis of transcranial and intraparenchymal light penetration in human cadaver brain tissue. *Lasers Surg. Med.* 2015;47:312–322. doi: 10.1002/lsm.22343. [[PubMed](#)] [[CrossRef](#)] [[Google Scholar](#)]

59. Fenghua T., Jenson V., Anh T., Qianqian F., Francisco G.-L. Effects of Wavelength on Transcranial Laser Stimulation: A Monte Carlo Simulation Study Based on Standard Brain Model (Conference Presentation); Proceedings of the Mechanisms of Photobiomodulation Therapy XV; San Francisco, CA, USA. 1 February 2020; San Francisco, CA, USA: Proc.SPIE; 2020. p. 112210B. [[Google Scholar](#)]

60. Henderson T.A., Morries L.D. Near-infrared photonic energy penetration: Can infrared phototherapy effectively reach the human brain? *Neuropsychiatr. Dis. Treat.* 2015;11:2191–2208. doi: 10.2147/NDT.S78182. [[PMC free article](#)] [[PubMed](#)] [[CrossRef](#)] [[Google Scholar](#)]

61. Tomas H., Donald D., Sean K., Peter J., Lars H. The Importance of Coherence in Phototherapy; Proceedings of the Mechanics for Low-Light Therapy; San Jose, CA, USA.

24 January 2009; San Jose, CA, USA: Proc.SPIE; 2009. p. 716507. [[Google Scholar](#)]

62. Litscher D., Litscher G. Laser therapy and dementia: A database analysis and future aspects on LED-based systems. *Int. J. Photoenergy.* 2014;2014:268354.

doi: 10.1155/2014/268354. [[CrossRef](#)] [[Google Scholar](#)]

63. Hamblin M.R. Shining light on the head: Photobiomodulation for brain disorders. *BBA Clin.*

2016;6:113–124. doi: 10.1016/j.bbaci.2016.09.002. [[PMC free article](#)] [[PubMed](#)] [[CrossRef](#)] [[Google Scholar](#)]

64. Freitas L.F.d., Hamblin M.R. Proposed mechanisms of photobiomodulation or low-level light therapy. *IEEE J. Sel. Top. Quant.* 2016;22:348–364.

doi: 10.1109/JSTQE.2016.2561201. [[PMC free article](#)] [[PubMed](#)] [[CrossRef](#)] [[Google Scholar](#)]

65. Liang H.L., Whelan H.T., Eells J.T., Meng H., Buchmann E., Lerch-Gaggl A., Wong-Riley M. Photobiomodulation partially rescues visual cortical neurons from cyanide-induced apoptosis. *Neuroscience.* 2006;139:639–649.

doi: 10.1016/j.neuroscience.2005.12.047. [[PubMed](#)] [[CrossRef](#)] [[Google Scholar](#)]

66. de Brito Sousa K., Rodrigues M.F.S.D., de Souza Santos D., Mesquita-Ferrari R.A., Nunes F.D., de Fátima Teixeira da Silva D., Bussadori S.K., Fernandes K.P.S. Differential expression of inflammatory and anti-inflammatory mediators by M1 and M2 macrophages after photobiomodulation with red or infrared lasers. *Lasers Med. Sci.* 2020;35:337–343.

doi: 10.1007/s10103-019-02817-1. [PubMed] [CrossRef] [Google Scholar]

67. Xuan W., Agrawal T., Huang L., Gupta G.K., Hamblin M.R. Low-level laser therapy for traumatic brain injury in mice increases brain derived neurotrophic factor (BDNF) and synaptogenesis. *J. Biophotonics*. 2015;8:502–511.

doi: 10.1002/jbio.201400069. [PMC free article] [PubMed] [CrossRef] [Google Scholar]

68. Weijun X., Fatma V., Liyi H., Michael R.H. Transcranial low-level laser therapy enhances learning, memory, and neuroprogenitor cells after traumatic brain injury in mice. *J. Biomed. Opt.* 2014;19:108003. [PMC free article] [PubMed] [Google Scholar]

69. Meng C., He Z., Da X. Low-level laser therapy rescues dendrite atrophy via upregulating BDNF expression: Implications for Alzheimer's disease. *J. Neurosci.* 2013;33:13505–13517. doi: 10.1523/JNEUROSCI.0918-13.2013. [PMC free article] [PubMed] [CrossRef] [Google Scholar]

70. Song S., Zhou F., Chen W.R. Low-level laser therapy regulates microglial function through Src-mediated signaling pathways: Implications for neurodegenerative diseases. *J. Neuroinflammation*. 2012;9:219. doi: 10.1186/1742-2094-9-219. [PMC free article] [PubMed] [CrossRef] [Google Scholar]

71. Tao L., Liu Q., Zhang F., Fu Y., Zhu X., Weng X., Han H., Huang Y., Suo Y., Chen L., et al. Microglia modulation with

1070-nm light attenuates A β burden and cognitive impairment in Alzheimer's disease mouse model. *Light Sci. Appl.* 2021;10:179. doi: 10.1038/s41377-021-00617-3. [[PMC free article](#)] [[PubMed](#)] [[CrossRef](#)] [[Google Scholar](#)]

72. O'Brien R.J., Wong P.C. Amyloid precursor protein processing and Alzheimer's disease. *Annu. Rev. Neurosci.* 2011;34:185–204. doi: 10.1146/annurev-neuro-061010-113613. [[PMC free article](#)] [[PubMed](#)] [[CrossRef](#)] [[Google Scholar](#)]

73. Lichtenthaler S.F., Haass C. Amyloid at the cutting edge: Activation of α -secretase prevents amyloidogenesis in an Alzheimer disease mouse model. *J. Clin. Invest.* 2004;113:1384–1387. doi: 10.1172/JCI21746. [[PMC free article](#)] [[PubMed](#)] [[CrossRef](#)] [[Google Scholar](#)]

74. Cho G.M., Lee S.-Y., Park J.H., Kim M.J., Park K.-J., Choi B.T., Shin Y.-I., Kim N.G., Shin H.K. Photobiomodulation using a low-level light-emitting diode improves cognitive dysfunction in the 5XFAD mouse model of Alzheimer's disease. *J. Gerontol. A Biol. Sci. Med. Sci.* 2020;75:631–639. doi: 10.1093/gerona/gly240. [[PubMed](#)] [[CrossRef](#)] [[Google Scholar](#)]

75. Sommer A.P., Bieschke J., Friedrich R.P., Zhu D., Wanker E.E., Fecht H.J., Mereles D., Hunstein W. 670 nm laser light and EGCG complementarily reduce amyloid- β aggregates in human neuroblastoma cells: Basis for treatment of Alzheimer's disease? *Photomed. Laser Surg.* 2011;30:54–60. doi: 10.1089/pho.2011.3073. [[PubMed](#)] [[CrossRef](#)] [[Google Scholar](#)]

[Scholar](#)

76. Allen N.J., Barres B.A. Glia—More than just brain glue. *Nature*. 2009;457:675–677. doi: 10.1038/457675a. [\[PubMed\]](#) [\[CrossRef\]](#) [\[Google Scholar\]](#)
77. von Leden R.E., Cooney S.J., Ferrara T.M., Zhao Y., Dalgard C.L., Anders J.J., Byrnes K.R. 808 nm wavelength light induces a dose-dependent alteration in microglial polarization and resultant microglial induced neurite growth. *Lasers Surg. Med.* 2013;45:253–263. doi: 10.1002/lsm.22133. [\[PubMed\]](#) [\[CrossRef\]](#) [\[Google Scholar\]](#)
78. Tsay H.-J., Huang Y.-C., Huang F.-L., Chen C.-P., Tsai Y.-C., Wang Y.-H., Wu M.-F., Chiang F.-Y., Shiao Y.-J. Amyloid β peptide-mediated neurotoxicity is attenuated by the proliferating microglia more potently than by the quiescent phenotype. *J. Biomed. Sci.* 2013;20:78. doi: 10.1186/1423-0127-20-78. [\[PMC free article\]](#) [\[PubMed\]](#) [\[CrossRef\]](#) [\[Google Scholar\]](#)
79. Oron A., Oron U. Low-level laser therapy to the bone marrow ameliorates neurodegenerative disease progression in a mouse model of Alzheimer's disease: A minireview. *Photomed. Laser Surg.* 2016;34:627. doi: 10.1089/pho.2015.4072. [\[PubMed\]](#) [\[CrossRef\]](#) [\[Google Scholar\]](#)
80. Mikita J., Dubourdieu-Cassagno N., Deloire M.S.A., Vekris A., Biran M., Raffard G., Brochet B., Canron M.-H., Franconi J.-M., Boiziau C., et al. Altered M1/M2 activation

patterns of monocytes in severe relapsing experimental rat model of multiple sclerosis. Amelioration of clinical status by M2 activated monocyte administration. *Mult. Scler.* 2010;17:2–15. doi: 10.1177/1352458510379243. [[PubMed](#)] [[CrossRef](#)] [[Google Scholar](#)]

81. Stepanov Y.V., Golovynska I., Zhang R., Golovynskyi S., Stepanova L.I., Gorbach O., Dovbyntchuk T., Garmanchuk L.V., Ohulchanskyy T.Y., Qu J. Near-infrared light reduces β -amyloid-stimulated microglial toxicity and enhances survival of neurons: Mechanisms of light therapy for Alzheimer's disease. *Alzheimers Res. Ther.* 2022;14:84.

doi: 10.1186/s13195-022-01022-7. [[PMC free article](#)] [[PubMed](#)] [[CrossRef](#)] [[Google Scholar](#)]

82. Sofroniew M.V. Multiple roles for astrocytes as effectors of cytokines and inflammatory mediators. *Neuroscientist.* 2013;20:160–172. doi: 10.1177/1073858413504466.

[[PubMed](#)] [[CrossRef](#)] [[Google Scholar](#)]

83. Jo S., Yarishkin O., Hwang Y.J., Chun Y.E., Park M., Woo D.H., Bae J.Y., Kim T., Lee J., Chun H., et al. GABA from reactive astrocytes impairs memory in mouse models of Alzheimer's disease. *Nat. Med.* 2014;20:886–896.

doi: 10.1038/nm.3639. [[PMC free article](#)] [[PubMed](#)] [[CrossRef](#)] [[Google Scholar](#)]

84. Yang X., Askarova S., Sheng W., Chen J.K., Sun A.Y., Sun G.Y., Yao G., Lee J.C.M. Low energy laser light (632.8 nm) suppresses amyloid- β peptide-induced oxidative and inflammatory responses in astrocytes. *Neuroscience.*

2010;171:859–868. doi: 10.1016/j.neuroscience.2010.09.025.

[[PMC free article](#)] [[PubMed](#)] [[CrossRef](#)] [[Google Scholar](#)]

85. Silveira P.C.L., Ferreira G.K., Zaccaron R.P., Glaser V., Remor A.P., Mendes C., Pinho R.A., Latini A. Effects of photobiomodulation on mitochondria of brain, muscle, and C6 astroglioma cells. *Med. Eng. Phys.* 2019;71:108–113. doi: 10.1016/j.medengphy.2019.05.008. [[PubMed](#)] [[CrossRef](#)] [[Google Scholar](#)]

86. Yoon S.-R., Hong N., Lee M.-Y., Ahn J.-C. Photobiomodulation with a 660-nanometer light-emitting diode promotes cell proliferation in astrocyte culture. *Cells.* 2021;10:1664. doi: 10.3390/cells10071664. [[PMC free article](#)] [[PubMed](#)] [[CrossRef](#)] [[Google Scholar](#)]

87. Han M., Wang Q., Wang X., Zeng Y., Huang Y., Meng Q., Zhang J., Wei X. Near infra-red light treatment of Alzheimer's disease. *J. Innov. Opt. Heal. Sci.* 2017;11:1750012. doi: 10.1142/S1793545817500122. [[CrossRef](#)] [[Google Scholar](#)]

88. Iaccarino H.F., Singer A.C., Martorell A.J., Rudenko A., Fan G., Gillingham T.Z., Mathys H., Seo J., Kritskiy O., Abdurrob F. Gamma frequency entrainment attenuates amyloid load and modifies microglia. *Nature.* 2016;540:230–235. doi: 10.1038/nature20587. [[PMC free article](#)] [[PubMed](#)] [[CrossRef](#)] [[Google Scholar](#)]

89. Martorell A.J., Paulson A.L., Suk H.-J., Abdurrob F., Drummond G.T., Guan W., Young J.Z., Kim D.N.-W., Kritskiy O., Barker S.J., et al. Multi-sensory gamma stimulation

ameliorates Alzheimer's-associated pathology and improves cognition. *Cell*. 2019;177:256–271.

doi: 10.1016/j.cell.2019.02.014. [[PMC free article](#)] [[PubMed](#)] [[CrossRef](#)] [[Google Scholar](#)]

90. Adaikkan C., Middleton S.J., Marco A., Pao P.-C., Mathys H., Kim D.N.-W., Gao F., Young J.Z., Suk H.-J., Boyden E.S., et al. Gamma entrainment binds higher-order brain regions and offers neuroprotection. *Neuron*. 2019;102:929–943.

doi: 10.1016/j.neuron.2019.04.011. [[PMC free article](#)] [[PubMed](#)] [[CrossRef](#)] [[Google Scholar](#)]

91. Yang L., Wu C., Parker E., Li Y., Dong Y., Tucker L., Brann D.W., Lin H.W., Zhang Q. Non-invasive photobiomodulation treatment in an Alzheimer Disease-like transgenic rat model. *Theranostics*. 2022;12:2205–2231. doi: 10.7150/thno.70756.

[[PMC free article](#)] [[PubMed](#)] [[CrossRef](#)] [[Google Scholar](#)]

92. Bonner J.F., Haas C.J., Fischer I. *Neuronal Cell Culture: Methods and Protocols*. Springer; Berlin/Heidelberg, Germany: 2013. Preparation of neural stem cells and progenitors: Neuronal production and grafting applications; pp. 65–88. [[PubMed](#)] [[Google Scholar](#)]

93. Wu X., Shen Q., Zhang Z., Zhang D., Gu Y., Xing D. Photoactivation of TGF β /SMAD signaling pathway ameliorates adult hippocampal neurogenesis in Alzheimer's disease model. *Stem Cell Res. Ther.* 2021;12:345.

doi: 10.1186/s13287-021-02399-2. [[PMC free article](#)] [[PubMed](#)] [[CrossRef](#)] [[Google Scholar](#)]

94. Zuo Z., Qi F., Yang J., Wang X., Wu Y., Wen Y., Yuan Q.,

Zou J., Guo K., Yao Z.B. Immunization with Bacillus Calmette-Guérin (BCG) alleviates neuroinflammation and cognitive deficits in APP/PS1 mice via the recruitment of inflammation-resolving monocytes to the brain. *Neurobiol. Dis.* 2017;101:27–39. doi: 10.1016/j.nbd.2017.02.001.

[[PubMed](#)] [[CrossRef](#)] [[Google Scholar](#)]

95. Baruch K., Rosenzweig N., Kertser A., Deczkowska A., Sharif A.M., Spinrad A., Tsitsou-Kampeli A., Sarel A., Cahalon L., Schwartz M. Breaking immune tolerance by targeting Foxp3(+) regulatory T cells mitigates Alzheimer's disease pathology. *Nat. Commun.* 2015;6:7967.

doi: 10.1038/ncomms8967. [[PMC free article](#)] [[PubMed](#)]
[[CrossRef](#)] [[Google Scholar](#)]

96. Dansokho C., Ait Ahmed D., Aid S., Toly-Ndour C., Chaigneau T., Calle V., Cagnard N., Holzenberger M., Piaggio E., Aucouturier P., et al. Regulatory T cells delay disease progression in Alzheimer-like pathology. Pt 4 *Brain*.

2016;139:1237–1251. doi: 10.1093/brain/awv408. [[PubMed](#)]
[[CrossRef](#)] [[Google Scholar](#)]

97. Arcuri C., Mecca C., Giambanco I., Donato R. Parenchymal and non-parenchymal immune cells in the brain: A critical role in regulating CNS functions. *Int. J. Dev. Neurosci.* 2019;77:26–38.

doi: 10.1016/j.ijdevneu.2019.04.005. [[PubMed](#)] [[CrossRef](#)]
[[Google Scholar](#)]

98. Ziv Y., Ron N., Butovsky O., Landa G., Sudai E., Greenberg N., Cohen H., Kipnis J., Schwartz M. Immune cells

contribute to the maintenance of neurogenesis and spatial learning abilities in adulthood. *Nat. Neurosci.* 2006;9:268–275. doi: 10.1038/nn1629. [PubMed] [CrossRef] [Google Scholar]

99. McGowan P.O., Hope T.A., Meck W.H., Kelsoe G., Williams C.L. Impaired social recognition memory in recombination activating gene 1-deficient mice. *Brain Res.* 2011;1383:187–195. doi: 10.1016/j.brainres.2011.02.054.

[PMC free article] [PubMed] [CrossRef] [Google Scholar]

100. Wu X., Shen Q., Chang H., Li J., Xing D. Promoted CD4+ T cell-derived IFN- γ /IL-10 by photobiomodulation therapy modulates neurogenesis to ameliorate cognitive deficits in APP/PS1 and 3xTg-AD mice. *J. Neuroinflamm.* 2022;19:253. doi: 10.1186/s12974-022-02617-5. [PMC free article]

[PubMed] [CrossRef] [Google Scholar]

101. Maksimovich I.V. Stimulation of cerebral angiogenesis and neurogenesis with transcatheter intracerebral laser photobiomodulation therapy during dementia in patients with Alzheimer's and Binswanger's disease. *Alzheimers Dement.* 2021;17:e054945. doi: 10.1002/alz.054945.

[CrossRef] [Google Scholar]

102. Shefer G., Partridge T.A., Heslop L., Gross J.G., Oron U., Halevy O. Low-energy laser irradiation promotes the survival and cell cycle entry of skeletal muscle satellite cells. Pt 7. *J. Cell Sci.* 2002;115:1461–1469. doi: 10.1242/jcs.115.7.1461.

[PubMed] [CrossRef] [Google Scholar]

103. Duan R., Zhu L., Liu T.C.-Y., Li Y., Liu J., Jiao J., Xu X.,

Yao L., Liu S. Light emitting diode irradiation protect against the amyloid beta 25–35 induced apoptosis of PC12 cell in vitro. *Lasers Surg. Med.* 2003;33:199–203.
doi: 10.1002/lsm.10216. [[PubMed](#)] [[CrossRef](#)] [[Google Scholar](#)]

104. Ghanbari A., Ghareghani M., Zibara K., Delaviz H., Ebadi E., Jahantab M.H. Light-Emitting Diode (LED) therapy improves occipital cortex damage by decreasing apoptosis and increasing BDNF-expressing cells in methanol-induced toxicity in rats. *Biomed. Pharmacother.* 2017;89:1320–1330.
doi: 10.1016/j.biopha.2017.03.024. [[PubMed](#)] [[CrossRef](#)] [[Google Scholar](#)]

105. Yu Z., Li Z., Liu N., Jizhang Y., McCarthy T.J., Tedford C.E., Lo E.H., Wang X. Near infrared radiation protects against oxygen-glucose deprivation-induced neurotoxicity by down-regulating neuronal nitric oxide synthase (nNOS) activity in vitro. *Metab. Brain Dis.* 2015;30:829–837.
doi: 10.1007/s11011-015-9663-3. [[PubMed](#)] [[CrossRef](#)] [[Google Scholar](#)]

106. Gavish L., Asher Y., Becker Y., Kleinman Y. Low level laser irradiation stimulates mitochondrial membrane potential and disperses subnuclear promyelocytic leukemia protein. *Lasers Surg. Med.* 2004;35:369–376.
doi: 10.1002/lsm.20108. [[PubMed](#)] [[CrossRef](#)] [[Google Scholar](#)]

107. Zhang L., Xing D., Zhu D., Chen Q. Low-power laser irradiation inhibiting A β 25–35-induced PC12 cell apoptosis

via PKC activation. *Cell. Physiol. Biochem.* 2008;22:215–222. doi: 10.1159/000149799. [[PubMed](#)] [[CrossRef](#)] [[Google Scholar](#)]

108. Zhang L., Zhang Y., Xing D. LPLI inhibits apoptosis upstream of Bax translocation via a GSK-3 β -inactivation mechanism. *J. Cell. Physiol.* 2010;224:218–228. doi: 10.1002/jcp.22123. [[PubMed](#)] [[CrossRef](#)] [[Google Scholar](#)]

109. Uozumi Y., Nawashiro H., Sato S., Kawauchi S., Shima K., Kikuchi M. Targeted increase in cerebral blood flow by transcranial near-infrared laser irradiation. *Lasers Surg. Med.* 2010;42:566–576. doi: 10.1002/lsm.20938. [[PubMed](#)] [[CrossRef](#)] [[Google Scholar](#)]

110. Lee H.I., Lee S.-W., Kim S.Y., Kim N.G., Park K.-J., Choi B.T., Shin Y.-I., Shin H.K. Pretreatment with light-emitting diode therapy reduces ischemic brain injury in mice through endothelial nitric oxide synthase-dependent mechanisms. *Biochem. Biophys. Res. Commun.* 2017;486:945–950. doi: 10.1016/j.bbrc.2017.03.131. [[PubMed](#)] [[CrossRef](#)] [[Google Scholar](#)]

111. Hennessy M., Hamblin M.R. Photobiomodulation and the brain: A new paradigm. *J. Opt.* 2017;19:013003. doi: 10.1088/2040-8986/19/1/013003. [[PMC free article](#)] [[PubMed](#)] [[CrossRef](#)] [[Google Scholar](#)]

112. De Taboada L., Yu J., El-Amouri S., Gattoni-Celli S., Richieri S., McCarthy T., Streeter J., Kindy M.S. Transcranial laser therapy attenuates amyloid-beta peptide

neuropathology in amyloid-beta protein precursor transgenic mice. *J. Alzheimers Dis.* 2011;23:521–535. doi: 10.3233/JAD-2010-100894. [PubMed] [CrossRef] [Google Scholar]

113. Telerman A., Lapter S., Sharabi A., Zinger H., Mozes E. Induction of hippocampal neurogenesis by a tolerogenic peptide that ameliorates lupus manifestations. *J. Neuroimmunol.* 2011;232:151–157. doi: 10.1016/j.jneuroim.2010.11.001. [PubMed] [CrossRef] [Google Scholar]

114. Yan X., Liu J., Zhang Z., Li W., Sun S., Zhao J., Dong X., Qian J., Sun H. Low-level laser irradiation modulates brain-derived neurotrophic factor mRNA transcription through calcium-dependent activation of the ERK/CREB pathway. *Lasers Med. Sci.* 2017;32:169–180. doi: 10.1007/s10103-016-2099-0. [PubMed] [CrossRef] [Google Scholar]

115. Sutalangka C., Wattanathorn J., Muchimapura S., Thukham-mee W., Wannanon P., Tong-un T. Laser acupuncture improves memory impairment in an animal model of Alzheimer's disease. *J. Acupunct. Meridian.* 2013;6:247–251. doi: 10.1016/j.jams.2013.07.001. [PubMed] [CrossRef] [Google Scholar]

116. Chan D., Suk H.-J., Jackson B.L., Milman N., Stark D., Fernandez V., Banerjee A., Kitchener E., Klerman E.B., Boyden E.S., et al. Gamma frequency sensory stimulation prevents brain atrophy, improves sleep and memory in probable mild Alzheimer's patients. *Alzheimers Dement.*

2021;17:e054218. doi: 10.1002/alz.054218. [[CrossRef](#)]

[[Google Scholar](#)]

117. Wang Z., Zhang Y., Dong L., Zheng Z., Zhong D., Long X., Cai Q., Jian W., Zhang S., Wu W., et al. Effects of morning blue-green 500 nm light therapy on cognition and biomarkers in middle-aged and older adults with subjective cognitive decline and mild cognitive impairment: Study protocol for a randomized controlled trial. *J. Alzheimers Dis.* 2021;83:1521–1536. doi: 10.3233/JAD-201560. [[PubMed](#)]

[[CrossRef](#)] [[Google Scholar](#)]

118. Purushothuman S., Johnstone D.M., Nandasena C., Mitrofanis J., Stone J. Photobiomodulation with near infrared light mitigates Alzheimer's disease-related pathology in cerebral cortex—evidence from two transgenic mouse models. *Alzheimers Res. Ther.* 2014;6:2.

doi: 10.1186/alzrt232. [[PMC free article](#)] [[PubMed](#)]

[[CrossRef](#)] [[Google Scholar](#)]

119. Purushothuman S., Johnstone D.M., Nandasena C., van Eersel J., Ittner L.M., Mitrofanis J., Stone J. Near infrared light mitigates cerebellar pathology in transgenic mouse models of dementia. *Neurosci. Lett.* 2015;591:155–159.

doi: 10.1016/j.neulet.2015.02.037. [[PubMed](#)] [[CrossRef](#)]

[[Google Scholar](#)]

120. da Luz Eltchechem C., Salgado A.S.I., Zângaro R.A., da Silva Pereira M.C., Kerppers I.I., da Silva L.A., Parreira R.B. Transcranial LED therapy on amyloid- β toxin 25–35 in the hippocampal region of rats. *Lasers Med. Sci.* 2017;32:749–

756. doi: 10.1007/s10103-017-2156-3. [[PubMed](#)] [[CrossRef](#)]
[[Google Scholar](#)]

121. Comerota M.M., Krishnan B., Taglialatela G. Near infrared light decreases synaptic vulnerability to amyloid beta oligomers. *Sci. Rep.* 2017;7:15012.
doi: 10.1038/s41598-017-15357-x. [[PMC free article](#)]
[[PubMed](#)] [[CrossRef](#)] [[Google Scholar](#)]

122. Arakelyan H.S. Treatment of Alzheimer's disease with a combination of laser, magnetic field and chromo light (colour) therapies: A double-blind controlled trial based on a review and overview of the etiological pathophysiology of Alzheimer's disease. *Laser Ther.* 2005;14:19–28.

doi: 10.5978/isism.14.19. [[CrossRef](#)] [[Google Scholar](#)]

123. Shen Q., Liu L., Gu X., Xing D. Photobiomodulation suppresses JNK3 by activation of ERK/MKP7 to attenuate AMPA receptor endocytosis in Alzheimer's disease. *Aging Cell.* 2021;20:e13289. doi: 10.1111/acel.13289. [[PMC free article](#)]
[[PubMed](#)] [[CrossRef](#)] [[Google Scholar](#)]

124. Shen Q., Wu X., Zhang Z., Zhang D., Yang S., Xing D. Gamma frequency light flicker regulates amyloid precursor protein trafficking for reducing β-amyloid load in Alzheimer's disease model. *Aging Cell.* 2022;21:e13573.
doi: 10.1111/acel.13573. [[PMC free article](#)] [[PubMed](#)]
[[CrossRef](#)] [[Google Scholar](#)]

125. Chan A.S., Lee T.-I., Hamblin M.R., Cheung M.-c. Photobiomodulation enhances memory processing in older adults with mild cognitive impairment: A functional near-

- infrared spectroscopy study. *J. Alzheimers Dis.* 2021;83:1471–1480. doi: 10.3233/JAD-201600. [[PubMed](#)] [[CrossRef](#)] [[Google Scholar](#)]
126. Saltmarche A.E., Naeser M.A., Ho K.F., Hamblin M.R., Lim L. Significant improvement in cognition in mild to moderately severe dementia cases treated with transcranial plus intranasal photobiomodulation: Case series report. *Photomed. Laser Surg.* 2017;35:432–441. doi: 10.1089/pho.2016.4227. [[PMC free article](#)] [[PubMed](#)] [[CrossRef](#)] [[Google Scholar](#)]
127. Zomorrodi R., Saltmarche A.E., Loheswaran G., Ho K.F., Lim L. Complementary EEG evidence for a significantly improved Alzheimer's disease case after photoboomodulation treatment. *Alzheimers Dement.* 2017;13:621. doi: 10.1016/j.jalz.2017.06.691. [[CrossRef](#)] [[Google Scholar](#)]
128. Chao L.L. Effects of home photobiomodulation treatments on cognitive and behavioral function, cerebral perfusion, and resting-state functional connectivity in patients with dementia: A pilot trial. *Photobiomodul Photomed. Laser Surg.* 2019;37:133–141. doi: 10.1089/photob.2018.4555. [[PubMed](#)] [[CrossRef](#)] [[Google Scholar](#)]
129. Michalikova S., Ennaceur A., van Rensburg R., Chazot P.L. Emotional responses and memory performance of middle-aged CD1 mice in a 3D maze: Effects of low infrared light. *Neurobiol. Learn. Mem.* 2008;89:480–488.

doi: 10.1016/j.nlm.2007.07.014. [PubMed] [CrossRef] [Google Scholar]

130. Grillo S.L., Duggett N.A., Ennaceur A., Chazot P.L. Non-invasive infra-red therapy (1072nm) reduces β -amyloid protein levels in the brain of an Alzheimer's disease mouse model, TASTPM. *J. Photochem. Photobiol. B Biol.* 2013;123:13–22. doi: 10.1016/j.jphotobiol.2013.02.015.

[PubMed] [CrossRef] [Google Scholar]

131. Zinchenko E., Navolokin N., Shirokov A., Khlebtsov B., Dubrovsky A., Saranceva E., Abdurashitov A., Khorovodov A., Terskov A., Mamedova A., et al. Pilot study of transcranial photobiomodulation of lymphatic clearance of beta-amyloid from the mouse brain: Breakthrough strategies for non-pharmacologic therapy of Alzheimer's disease. *Biomed. Opt. Express.* 2019;10:4003–4017. doi: 10.1364/BOE.10.004003.

[PMC free article] [PubMed] [CrossRef] [Google Scholar]

132. Semyachkina-Glushkovskaya O., Penzel T., Blokhina I., Khorovodov A., Fedosov I., Yu T., Karandin G., Evsukova A., Elovenko D., Adushkina V., et al. Night photostimulation of clearance of beta-amyloid from mouse brain: New strategies in preventing Alzheimer's disease. *Cells.* 2021;10:3289.

doi: 10.3390/cells10123289. [PMC free article] [PubMed] [CrossRef] [Google Scholar]

133. Berman M.H., Halper J.P., Nichols T.W., Jarrett H., Lundy A., Huang J.H. Photobiomodulation with near infrared light helmet in a pilot, placebo controlled clinical trial in dementia patients testing memory and cognition. *J. Neurol. Neurosci.*

2017;8:176. doi: 10.21767/2171-6625.1000176. [[PMC free article](#)] [[PubMed](#)] [[CrossRef](#)] [[Google Scholar](#)]

134. Nizamutdinov D., Qi X., Berman M.H., Dougal G., Dayawansa S., Wu E., Yi S.S., Stevens A.B., Huang J.H. Transcranial near infrared light stimulations improve cognition in patients with dementia. *Aging Dis.* 2021;12:954–963. doi: 10.14336/AD.2021.0229. [[PMC free article](#)] [[PubMed](#)] [[CrossRef](#)] [[Google Scholar](#)]

135. Qi X., Nizamutdinov D., Berman M.H., Dougal G., Chazot P.L., Wu E., Stevens A.B., Yi S.S., Huang J.H. Gender differences of dementia in response to intensive self-administered transcranial and intraocular near-infrared stimulation. *Cureus.* 2021;13:e16188. doi: 10.7759/cureus.16188. [[PMC free article](#)] [[PubMed](#)] [[CrossRef](#)] [[Google Scholar](#)]

136. Qu X., Li L., Zhou X., Dong Q., Liu H., Liu H., Yang Q., Han Y., Niu H. Repeated transcranial photobiomodulation improves working memory of healthy older adults: Behavioral outcomes of poststimulation including a three-week follow-up. *Neurophotonics.* 2022;9:035005.

doi: 10.1111/1.NPh.9.3.035005. [[PMC free article](#)] [[PubMed](#)] [[CrossRef](#)] [[Google Scholar](#)]

137. Elsworth J.D. Parkinson's disease treatment: Past, present, and future. *J. Neural Transm.* 2020;127:785–791. doi: 10.1007/s00702-020-02167-1. [[PMC free article](#)] [[PubMed](#)] [[CrossRef](#)] [[Google Scholar](#)]

138. Toffoli M., Vieira S.R.L., Schapira A.H.V. Genetic causes

of PD: A pathway to disease modification.

Neuropharmacology. 2020;170:108022.

doi: 10.1016/j.neuropharm.2020.108022. [[PubMed](#)]

[[CrossRef](#)] [[Google Scholar](#)]

139. Zaia A., Maponi P., Zannotti M., Casoli T. Biocomplexity and fractality in the search of biomarkers of aging and pathology: Mitochondrial DNA profiling of Parkinson's disease. *Int. J. Mol. Sci.* 2020;21:1758.

doi: 10.3390/ijms21051758. [[PMC free article](#)] [[PubMed](#)]

[[CrossRef](#)] [[Google Scholar](#)]

140. Salehpour F., Hamblin M.R. Photobiomodulation for Parkinson's disease in animal models: A systematic review. *Biomolecules*. 2020;10:610. doi: 10.3390/biom10040610.

[[PMC free article](#)] [[PubMed](#)] [[CrossRef](#)] [[Google Scholar](#)]

141. Shaw V.E., Spana S., Ashkan K., Benabid A.-L., Stone J., Baker G.E., Mitrofanis J. Neuroprotection of midbrain dopaminergic cells in MPTP-treated mice after near-infrared light treatment. *J. Comp. Neurol.* 2010;518:25–40.

doi: 10.1002/cne.22207. [[PubMed](#)] [[CrossRef](#)] [[Google Scholar](#)]

142. Mitrofanis J. Why and how does light therapy offer neuroprotection in Parkinson's disease? *Neural Regen. Res.* 2017;12:574. doi: 10.4103/1673-5374.205092. [[PMC free article](#)] [[PubMed](#)] [[CrossRef](#)] [[Google Scholar](#)]

143. Eiden L.E., Weihe E. VMAT2: A dynamic regulator of brain monoaminergic neuronal function interacting with drugs of abuse. *Ann. N. Y. Acad. Sci.* 2011;1216:86–98.

doi: 10.1111/j.1749-6632.2010.05906.x. [[PMC free article](#)] [[PubMed](#)] [[CrossRef](#)] [[Google Scholar](#)]

144. El Massri N., Lemgruber A.P., Rowe I.J., Moro C., Torres N., Reinhart F., Chabrol C., Benabid A.-L., Mitrofanis J. Photobiomodulation-induced changes in a monkey model of Parkinson's disease: Changes in tyrosine hydroxylase cells and GDNF expression in the striatum. *Exp. Brain Res.* 2017;235:1861–1874. doi: 10.1007/s00221-017-4937-0.

[[PubMed](#)] [[CrossRef](#)] [[Google Scholar](#)]

145. Gu X., Liu L., Shen Q., Xing D. Photoactivation of ERK/CREB/VMAT2 pathway attenuates MPP+-induced neuronal injury in a cellular model of Parkinson's disease. *Cell. Signal.* 2017;37:103–114.

doi: 10.1016/j.cellsig.2017.06.007. [[PubMed](#)] [[CrossRef](#)] [[Google Scholar](#)]

146. Zhang Z., Lei L., Jiang X., Zhai S., Da X. The essential role of Drp1 and its regulation by S-nitrosylation of Parkin in dopaminergic neurodegeneration: Implications for Parkinson's disease. *Antioxid. Redox Signal.* 2016;25:609–622. doi: 10.1089/ars.2016.6634. [[PubMed](#)] [[CrossRef](#)] [[Google Scholar](#)]

147. Wang R., Dong Y., Lu Y., Zhang W., Brann D.W., Zhang Q. Photobiomodulation for global cerebral ischemia: Targeting mitochondrial dynamics and functions. *Mol. Neurobiol.* 2019;56:1852–1869. doi: 10.1007/s12035-018-1191-9. [[PMC free article](#)] [[PubMed](#)] [[CrossRef](#)] [[Google Scholar](#)]

148. Liang H.L., Whelan H.T., Eells J.T., Wong-Riley M.T.T.

Near-infrared light via light-emitting diode treatment is therapeutic against rotenone- and 1-methyl-4-phenylpyridinium ion-induced neurotoxicity. *Neuroscience*. 2008;153:963–974.

doi: 10.1016/j.neuroscience.2008.03.042. [[PMC free article](#)] [[PubMed](#)] [[CrossRef](#)] [[Google Scholar](#)]

149. Johnstone D.M., el Massri N., Moro C., Spana S., Wang X.S., Torres N., Chabrol C., De Jaeger X., Reinhart F., Purushothuman S., et al. Indirect application of near infrared light induces neuroprotection in a mouse model of parkinsonism—An abscopal neuroprotective effect. *Neuroscience*. 2014;274:93–101.

doi: 10.1016/j.neuroscience.2014.05.023. [[PubMed](#)] [[CrossRef](#)] [[Google Scholar](#)]

150. Moro C., Massri N.E., Torres N., Ratel D., De Jaeger X., Chabrol C., Perraut F., Bourgerette A., Berger M., Purushothuman S., et al. Photobiomodulation inside the brain: A novel method of applying near-infrared light intracranially and its impact on dopaminergic cell survival in MPTP-treated mice: Laboratory investigation. *J. Neurosurg*. 2014;120:670–683. doi: 10.3171/2013.9.JNS13423.

[[PubMed](#)] [[CrossRef](#)] [[Google Scholar](#)]

151. Wattanathorn J., Satalangka C. Laser acupuncture at HT7 acupoint improves cognitive deficit, neuronal loss, oxidative stress, and functions of cholinergic and dopaminergic systems in animal model of Parkinson's disease. *Evid. Based Complement. Alternat. Med.*

2014;2014:937601. doi: 10.1155/2014/937601. [[PMC free article](#)] [[PubMed](#)] [[CrossRef](#)] [[Google Scholar](#)]

152. Peoples C., Spana S., Ashkan K., Benabid A.L., Stone J., Baker G.E., Mitrofanis J. Photobiomodulation enhances nigral dopaminergic cell survival in a chronic MPTP mouse model of Parkinson's disease. *Park. Relat. Disord.* 2012;18:469–476. doi: 10.1016/j.parkreldis.2012.01.005. [[PubMed](#)] [[CrossRef](#)] [[Google Scholar](#)]

153. Moro C., Torres N., El Massri N., Ratel D., Johnstone D.M., Stone J., Mitrofanis J., Benabid A.L.

Photobiomodulation preserves behaviour and midbrain dopaminergic cells from MPTP toxicity: Evidence from two mouse strains. *BMC Neurosci.* 2013;14:40.

doi: 10.1186/1471-2202-14-40. [[PMC free article](#)] [[PubMed](#)] [[CrossRef](#)] [[Google Scholar](#)]

154. Darlot F., Moro C., El Massri N., Chabrol C., Johnstone D.M., Reinhart F., Agay D., Torres N., Bekha D., Auboiron V., et al. Near-infrared light is neuroprotective in a monkey model of Parkinson disease. *Ann. Neurol.* 2016;79:59–75. doi: 10.1002/ana.24542. [[PubMed](#)] [[CrossRef](#)] [[Google Scholar](#)]

155. El Massri N., Moro C., Torres N., Darlot F., Agay D., Chabrol C., Johnstone D.M., Stone J., Benabid A.-L., Mitrofanis J. Near-infrared light treatment reduces astrogliosis in MPTP-treated monkeys. *Exp. Brain Res.* 2016;234:3225–3232. doi: 10.1007/s00221-016-4720-7. [[PubMed](#)] [[CrossRef](#)] [[Google Scholar](#)]

156. Moro C., El Massri N., Darlot F., Torres N., Chabrol C., Agay D., Auboiroux V., Johnstone D.M., Stone J., Mitrofanis J., et al. Effects of a higher dose of near-infrared light on clinical signs and neuroprotection in a monkey model of Parkinson's disease. *Brain Res.* 2016;1648:19–26.

doi: 10.1016/j.brainres.2016.07.005. [PubMed] [CrossRef] [Google Scholar]

157. Reinhart F., Massri N.E., Chabrol C., Cretallaz C., Johnstone D.M., Torres N., Darlot F., Costecalde T., Stone J., Mitrofanis J., et al. Intracranial application of near-infrared light in a hemi-parkinsonian rat model: The impact on behavior and cell survival. *J. Neurosurg.* 2016;124:1829–1841. doi: 10.3171/2015.5.JNS15735. [PubMed] [CrossRef] [Google Scholar]

158. Reinhart F., El Massri N., Johnstone D.M., Stone J., Mitrofanis J., Benabid A.-L., Moro C. Near-infrared light (670 nm) reduces MPTP-induced parkinsonism within a broad therapeutic time window. *Exp. Brain Res.* 2016;234:1787–1794. doi: 10.1007/s00221-016-4578-8. [PubMed] [CrossRef] [Google Scholar]

159. Shaw V.E., Peoples C., Spana S., Ashkan K., Benabid A.-L., Stone J., Baker G.E., Mitrofanis J. Patterns of cell activity in the subthalamic region associated with the neuroprotective action of near-infrared light treatment in MPTP-treated mice. *Park. Dis.* 2012;2012:296875. doi: 10.1155/2012/296875. [PMC free article] [PubMed] [CrossRef] [Google Scholar]

160. Purushothuman S., Nandasena C., Johnstone D.M., Stone J., Mitrofanis J. The impact of near-infrared light on dopaminergic cell survival in a transgenic mouse model of parkinsonism. *Brain Res.* 2013;1535:61–70.
doi: 10.1016/j.brainres.2013.08.047. [PubMed] [CrossRef] [Google Scholar]
161. El Massri N., Johnstone D.M., Peoples C.L., Moro C., Reinhart F., Torres N., Stone J., Benabid A.-L., Mitrofanis J. The effect of different doses of near infrared light on dopaminergic cell survival and gliosis in MPTP-treated mice. *Int. J. Neurosci.* 2016;126:76–87.
doi: 10.3109/00207454.2014.994063. [PubMed] [CrossRef] [Google Scholar]
162. El Massri N., Cullen K.M., Stefani S., Moro C., Torres N., Benabid A.-L., Mitrofanis J. Evidence for encephalopsin immunoreactivity in interneurones and striosomes of the monkey striatum. *Exp. Brain Res.* 2018;236:955–961.
doi: 10.1007/s00221-018-5191-9. [PubMed] [CrossRef] [Google Scholar]
163. Kim B., Mitrofanis J., Stone J., Johnstone D.M. Remote tissue conditioning is neuroprotective against MPTP insult in mice. *IBRO Rep.* 2018;4:14–17.
doi: 10.1016/j.ibror.2018.01.001. [PMC free article] [PubMed] [CrossRef] [Google Scholar]
164. O'Brien J.A., Austin P.J. Effect of photobiomodulation in rescuing lipopolysaccharide-induced dopaminergic cell loss in the male Sprague–Dawley rat. *Biomolecules*. 2019;9:381.

doi: 10.3390/biom9080381. [[PMC free article](#)] [[PubMed](#)] [[CrossRef](#)] [[Google Scholar](#)]

165. San Miguel M., Martin K.L., Stone J., Johnstone D.M. Photobiomodulation mitigates cerebrovascular leakage induced by the Parkinsonian neurotoxin MPTP. *Biomolecules*. 2019;9:564. doi: 10.3390/biom9100564. [[PMC free article](#)] [[PubMed](#)] [[CrossRef](#)] [[Google Scholar](#)]

166. Ganeshan V., Skladnev N.V., Kim J.Y., Mitrofanis J., Stone J., Johnstone D.M. Pre-conditioning with remote photobiomodulation modulates the brain transcriptome and protects against MPTP insult in mice. *Neuroscience*. 2019;400:85–97. doi: 10.1016/j.neuroscience.2018.12.050. [[PubMed](#)] [[CrossRef](#)] [[Google Scholar](#)]

167. Oueslati A., Lovisa B., Perrin J., Wagnières G., van den Bergh H., Tardy Y., Lashuel H.A. Photobiomodulation suppresses alpha-synuclein-induced toxicity in an AAV-based rat genetic model of Parkinson's disease. *PLoS ONE*. 2015;10:e0140880. doi: 10.1371/journal.pone.0140880. [[PMC free article](#)] [[PubMed](#)] [[CrossRef](#)] [[Google Scholar](#)]

168. Reinhart F., Massri N.E., Darlot F., Torres N., Johnstone D.M., Chabrol C., Costecalde T., Stone J., Mitrofanis J., Benabid A.-L., et al. 810 nm near-infrared light offers neuroprotection and improves locomotor activity in PTP-treated mice. *Neurosci. Res.* 2015;92:86–90. doi: 10.1016/j.neures.2014.11.005. [[PubMed](#)] [[CrossRef](#)] [[Google Scholar](#)]

169. Vos M., Lovisa B., Geens A., Morais V.A., Wagnières G.,

van den Bergh H., Ginggen A., De Strooper B., Tardy Y., Verstreken P. Near-infrared 808 nm light boosts complex IV-dependent respiration and rescues a Parkinson-related pink1 model. *PLoS ONE*. 2013;8:e78562.

doi: 10.1371/journal.pone.0078562. [[PMC free article](#)]

[[PubMed](#)] [[CrossRef](#)] [[Google Scholar](#)]

170. Bullock-Saxton J., Lehn A., Laakso E.L. Exploring the effect of combined transcranial and intra-oral photobiomodulation therapy over a four-week period on physical and cognitive outcome measures for people with Parkinson's disease: A randomized double-blind placebo-controlled pilot study. *J. Alzheimers Dis.* 2021;83:1499–1512.

doi: 10.3233/JAD-210170. [[PubMed](#)] [[CrossRef](#)] [[Google Scholar](#)]

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